## SCHOOL OF LIFE SCIENCES JAWAHARLAL NEHRU UNIVERSITY

## $M.Sc-II^{nd}$ Semester Practical Teaching Programme, (2019-20)

Course Name	Course In-charge/Faculty	Experiments
Molecular	Prof. P.C. Rath	1- Preparation of LB medium, LB+ agar(1.5%)+ ampicillin
Biology	Prof. K. Natarajan	(100µg/ml) and LB+agar(1.5%)+nalidixic
		acid(15µg/ml) plates and other reagents.
	(27 classes)	2- Streaking of <i>E. coli</i> DH5α strain on LB+agar+ nalidixic
		acid plate.
		3- Preparation of competent E. coli DH5α cells.
		4- Transformation of <i>E. coli</i> DH5α cells with pBluescript
		plasmid DNA and recombinant DNA clones.
		5- Isolation of plasmid DNA from overnight culture of the
		transformed colony/ clone.
		6- Agarose gel electrophoresis of the plasmid DNA.
		7- Restriction enzyme digestion of the plasmid DNA and
		clone verification.
		8- Restriction enzyme digestion of the becteriophage λ
		DNA/ genome by Hind III and Hind III+EcoR I and preparation of the restriction map for the double digest.
		9- Preparation of a ball and stick model of B-DNA. Draw
		the chemical structures of A,T and G,C base pairs, show
		different groups/ bonds in the DNA and study the
		structural features.
		10- Blue/ white screening of the recombinant DNA clone in
		E.coli.
		11- Lecture on PCR; preparation of reagents and
		media; inoculate for genomic DNA isolation from
		yeast
		12- Genomic DNA isolation and quantitation on
		gel
		13- Primer designing; Set up PCR using serial dilutions of
		genomic/plasmid DNA template
		Run gel, analyze of gel results and discussion.
		14- Expression of recombinant protein in E. coli.
Cell Biology	Dr. Neelima Mondal	1- Cell plating and harvesting.
	(5classes)	2- Fixing of the cells.
		3- Propidium Iodide staining of the cells.
		4- To study the cell cycle/ different phase of cell cycle.
		5- Analysis by ModFitprogramme.
Immunology	Dr. NitiPuri	1- Cell Counting of immune cells.
	(5classes)	2- Immunophenotyping experiment/labeling/ running
		sample on Flowcytometry followed by analysis.
		3- To Study antigen induced activation of immune cells.
		4- Detection of cytokines/chemokines in activated immune
G 11 1	D 4 5	cell through ELISA.
Cellular	Dr. A. Bansal	1- Staining and counting parasitemia of the malaria
Parasitology	(5 days)	parasite, <i>Plasmodium falciparum</i> .
		2- SYBER Green I based fluorescence assay for testing
		anti-malarial compounds.
		3- Detection of <i>Leishmania</i> parasite infection by
		fluorescence microscopy.
		4- Immunofluorescence assay (IFA) for testing protein localization in malaria parasite.
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Plant	Prof. A. Pareek, (5 days)	1- Study of phototropism in presence and absence of NPA
Physiology	Prof. S. Chakraborty (5 days)	in monocot and dicot plants.
T injured by	and	2- Isolation of chlorophyll from monocot and dicot plants
	Prof. A. Nandi	under different concentration, condition (dark-light) of
	(4 days)	IAA and BAP.
		3- Study of root and shoot length of monocot and dicot
		seedlings under different concentration of IAA and BAP
		under conditions (light-dark).
		4- To study protein profiling from monocot and dicot
		plants.
		5- Viral infection assays through agrobacterium
		tumefaciens inoculation and effect on host physiology.
l		6- Virus induced gene silencing of phytoene desaturase in
		N. benthamiana.
		7- Estimation of chlorophyll fluorescence through Pulse
		Amplititude Modulator (PAM).
		8- Separation of chlorophyll a and b by TLC.
		9- To study expression pattern of Auxin by using DR5-GUS transgenic lines in Arabidopsis.
		10- Effect of Absicis acid and Jasmonic acid on seed
		germination.
		11- To study triple response in Arabidopsis.
		12- Observation of different stage of embryo development.
		13- Observe plant stomata.
Animal	Dr. S. K. Jha (2 days) and	1- Demonstration and application of rodent's stereotaxic
Physiology	Dr. A. Mondal (3 days)	instruments.
		2- Demonstration of recorded brain waves. Identification of
		vigilant states in the recorded brain waves.
		3- Identification of alpha, beta, delta, theta and gamma brain
		waves in recorded the EEG.
		4- Study of squamous epithelial cells.
		5- Measurement of arterial blood pressure.
		6- Measurement of physical fitness Index (PFI) by Harvard
		step test.
		7- Effect of exercise on blood pressure & heart rate.
		8- Determination of blood group.
		<ul> <li>Study of haemin crystal.</li> </ul>
		<ul> <li>Preparation &amp; staining of human blood film.</li> </ul>
		9- TC & DC of RBC & WBC.
Microbial	Prof. Arun S. Kharat	1- Determination of Minimum Inhibitory Concentration.
growth and	(9 days)	2- Biofilm Demonstration and Quantification.
management		3- Bacterial Adherence and Invasion to Human Cell Line.
study		4- Demonstration of Antimicrobial activity from
		Medicinal Plant(s).

Coordinator